

Lack of Inhibitory Effect of SCH-39304 (An Antifungal Agent) on the Antiviral Efficacy of AZT Against HIV-1 *In Vitro*. J. B. Kahlon<sup>1</sup>, W. M. Shannon<sup>1</sup>, A. D. Brazier<sup>1</sup>, G. H. Miller<sup>2</sup>, J. Schwartz<sup>3</sup> and R. J. Whitley<sup>3</sup>. Southern Research Institute, Birmingham, AL 35255 USA<sup>1</sup>, Schering-Plough Corporation, Bloomfield, New Jersey 07003 USA<sup>2</sup>, and University of Alabama in Birmingham, Birmingham, AL 35205 USA<sup>3</sup>.

An anti-fungal agent for administration to AIDS patients and coded as SCH-39304 was tested for *in vitro* antiviral activity against the human immunodeficiency virus (HIV-1) in parallel with AZT using MT-2 cells (Harada *et al*, Science 229:563-566). Cytopathology inhibition data demonstrated that the drug has no activity against HIV-1. *In vitro* data were also generated in order to demonstrate a lack of enhancement of HIV-1 replication. Target cells (MT-2) were infected with varying multiplicities of infection of HIV-1 obtained from a standard stock preparation of the virus. A standard non-cytotoxic concentration of each drug (SCH-39304 = 100 µg/ml and AZT = 0.32 µg/ml) was used to measure enhancement of HIV-1 cytopathology in the target cells infected at varying multiplicities of infection. End-points were measured as percent viable cells, read as optical density values with the use of a tetrazolium salt (MTT) used for measuring cell viabilities. A combination of SCH-39304 with AZT was studied *in vitro* to measure antagonism or synergism between the two compounds. Again a non-cytotoxic concentration (100 µg/ml) of SCH-39304 was tested in combination with six half log<sub>10</sub> concentrations of AZT ranging from 0.032 µg/ml to 10 µg/ml in a viral cytopathology inhibition assay. Results demonstrated no antagonism between the two compounds at the tested concentrations. In an independent experiment, the effect of AZT and SCH-39304 combinations was studied for their potential ability to induce latent HIV-1 infections. U1 cells, a subclone of U937 cells, chronically infected with HIV-1 (Folks, T. M. *et al*, Science 238:800-802, 1987) were plated in the log phase of replication and treated with various concentrations of the two compounds. At day 3 post-treatment the cells were processed for hybridization and probed for replicating mRNA using a probe specific for HIV-1 pol gene (Gene-Trak Systems). Hybridizations were performed according to manufacturers instructions. Results show no detectable induction of viral replication in the treated samples.

Effect of Natural Products of Various Microbial Fermentations from Schering-Plough Corporation on the Replication of HIV-1 as Measured by CPE-Inhibition Assays in Infected Cells. P. Daniels<sup>1</sup>, S. Watkins<sup>1</sup>, J. Kahlon<sup>1</sup>, R. Hart<sup>2</sup>, A. C. Horan<sup>2</sup>, V. Gullo<sup>2</sup>, J. Schwartz<sup>3</sup>, R. J. Whitley<sup>3</sup>, and W. M. Shannon<sup>1</sup>. Southern Research Institute, Birmingham, AL 35255 USA<sup>1</sup>, Schering-Plough Corporation, Bloomfield, New Jersey 07003 USA<sup>2</sup>, and University of Alabama in Birmingham, Birmingham, AL 35205 USA<sup>3</sup>.

A total of 2,000 samples, products of various microbial fermentations, were screened for antiviral activity against the human immunodeficiency virus (HIV-1). Samples were tested according to our standard operating procedure (SOP) designed and implemented by scientists at Southern Research Institute and Schering-Plough Corporation. According to this SOP, each sample was tested at a single concentration (1:20 dilution of the submitted broth), in duplicate on HIV-1 infected MT-2 cells to measure antiviral efficacy and at the same concentration on uninfected MT-2 cells to measure the cytotoxic effect of the product. Assays were set up in 96-well plates, 18 products were tested per plate and three replicates of virus control cultures (untreated, infected cells) and three replicates of cell control cultures (untreated, uninfected cells) were used. A panel of control cultures (AZT and ddC at various concentrations; alone and in combination with selected fermentation products) were set up with each test (196 samples per test). Assay end-points were read at Day 7 post-infection by MTT, a formazan-activating tetrazolium salt used to measure cell viabilities (Tada, *et al*, J. Immunol Methods, 93:157-165). Criteria for selection of active compounds included arbitrary limits of 49% or less viability of test sample = inactive; 50-74 percent viability = marginally active; and 75-100% viability = active. Active and marginally active compounds were re-tested to confirm activity. Compounds meeting criteria of active are undergoing further *in vitro* activity and cytotoxicity testing.